

Glutamate-agonist-evoked taurine release from the adult and developing mouse hippocampus in cell-damaging conditions

P. Saransaari¹ and S. S. Oja^{1,2}

¹Tampere Brain Research Center, University of Tampere Medical School, and ²Department of Clinical Physiology, Tampere University Hospital, Tampere, Finland

Accepted June 17, 1997

Summary. Taurine is a neuromodulator and osmoregulator in the central nervous system, also protecting neural cells against excitotoxicity. The effects of the ionotropic glutamate receptor agonists N-methyl-D-aspartate (NMDA), kainate and 2-amino-3-hydroxy-5-methyl-4-imidazolepropionate (AMPA) on [3H]taurine release from hippocampal slices from 3-month-old and 7-day-old mice were studied in cell-damaging conditions. Neural cell injury was induced by superfusing the slices in hypoxic, hypoglycemic and ischemic conditions and by exposing them to metabolic poisons, free radicals and oxidative stress. The release of taurine was greatly enhanced in these conditions at both ages, except in oxidative stress. In normal conditions the three glutamate agonists potentiated taurine release in the immature hippocampus in a receptor-mediated manner, but kainate receptors did not participate in the regulation in the adults. The ability of the agonists to evoke taurine release varied in the cell-damaging conditions, but the glutamate-receptor-activated release was generally operating in the immature hippocampus. This glutamate-receptor-evoked massive release of taurine could have significant neuroprotective effects, particularly in the developing hippocampus, countering the harmful actions of the simultaneously liberated excitatory amino acids.

Keywords: Amino acids – Taurine release – Glutamate agonists – Cell damage – Tissue slices – Hippocampus

Introduction

Massive release of excitatory amino acids from neural structures during hypoxia and ischemia has been observed both in vitro (Pellegrini-Giampietro et al., 1990; Collard and Menon-Johansson, 1993; O'Regan et al., 1995) and in vivo (Benveniste et al., 1984; Hagberg et al., 1985; Globus et al., 1988). Excitatory amino acids are neurotoxic in excess and the resulting overstimulation of their receptors contributes to neuronal death during cerebral

ischemia (see Rothman and Olney, 1988; Szatkowski and Attwell, 1994). The excitatory amino acids are partly released as a result of the action of oxygenderived free radicals formed in hypoxic brain tissue (Pellegrini-Giampietro et 1988). Free radicals and excitatory amino acids cooperate in ischemia-induced neuronal damage (Pellegrini-Giampietro et al., 1990; Coyle and Puttfarcken, 1993). Of different brain areas, the hippocampus is particularly vulnerable to this kind of ischemic injury and subsequent cell death (Pulsinelli et al., 1992). It is particularly sensitive to excitotoxic agents in the newborn (Cook and Crutcher, 1986) but fairly well spared in hypoxia or ischemia (Cherubini et al., 1989; Ikonomidou et al., 1989; Ferriero et al., 1990). Since the most excitatory innervation in the hippocampus is glutamatergic, this vulnerability probably results from excitotoxicity mediated via glutamate receptors of the N-methyl-D-aspartate (NMDA) type (Zorumski and Olney, 1993), although the non-NMDA, kainate and 2amino-3-hydroxy-5-methyl-4-imidazolepropionate (AMPA), receptors could also be involved (Choi and Rothman, 1990; Obrenovitch and Urenjak, 1997).

The inhibitory amino acid taurine has been held to have a special role in immature brain tissue (Oja and Kontro, 1983; Kontro and Oja, 1987). It appears to be essential for the development and survival of neural cells (see Huxtable, 1992; Sturman, 1993). Taurine also protects these cells against excitotoxicity induced by excitatory amino acids (French et al., 1986; Trenkner, 1990; Tang et al., 1996) and prevents harmful metabolic events induced by ischemia or hypoxia (Schurr et al., 1987). Furthermore, taurine-containing neurons are relatively inert in cerebral ischemia induced by 4-vessel occlusion (Wu et al., 1994). Taurine has long been known to ameliorate symptoms in epilepsy (Oja and Kontro, 1983). These neuroprotective effects may be related to the functions of taurine as a neuromodulator (Saransaari and Oja, 1992), osmoregulator, antioxidant and regulator of calcium ion movements (Oja and Kontro, 1983; Huxtable, 1992; Oja and Saransaari, 1996).

Taurine abounds in the hippocampus (Lombardini, 1976; Kontro et al., 1980) and taurine-like immunoreactivity is located in hippocampal interneurons, pyramidal neurons and dentate granule cells (Magnusson et al., 1989). Taurine inhibits the firing of hippocampal pyramidal neurons by increasing chloride conductance and causing hyperpolarization (Taber et al., 1986). The enzyme synthesizing taurine, cysteine sulphinate decarboxylase, has also been identified in pyramidal basket interneurons (Taber et al., 1986). Hippocampal taurine release is markedly enhanced in cell-damaging conditions such as hypoxia, hypoglycemia, ischemia and exposure to free radicals and oxidative stress (Saransaari and Oja, 1996; 1997a). It has also been demonstrated that the ionotropic glutamate receptor agonists NMDA, kainate and AMPA, evoke taurine release in both developing and adult hippocampus (Magnusson et al., 1991; Saransaari and Oja, 1994). We have now investigated how these ionotropic glutamate agonists evoke the release of preloaded [3H]taurine from hippocampal slices from developing and adult mice in the above cell-damaging conditions.

Material and methods

Material

NMRI mice of both sexes aged 7 days and 3 months (adults), were used throughout. [1,2-3H]Taurine (specific activity 1.07 PBq/mol) was obtained from Amersham International, Bristol, UK. Dizocilpine (MK-801) was a gift from Merck, Sharp & Dohme (Rahway, NJ).

Efflux experiments

Slices 0.4 mm thick weighing 15–20 mg were prepared from the hippocampi with a Stadie-Riggs tissue slicer and used immediately in efflux experiments. The slices were preloaded for 30 min with $10\mu M$ (50 MBq/l) [³H]taurine in preoxygenated Krebs-Ringer-Hepes-glucose medium under O_2 and then superfused as described in Kontro and Oja (1987). The medium was pooled during the first 20 min of superfusion and thereafter 2-min fractions (0.5 ml) were collected. The glutamate receptor agonists and antagonists were added at 30 min. After superfusion the slices were weighed, homogenized in ice-cold 5% (w/v) trichloracetic acid solution and centrifuged, and the clear supernatants used for scintillation counting. The effluent samples were likewise counted for radioactivity.

Estimation of efflux rate constants

The release of labeled taurine from the slices was plotted as a function of time on the basis of the radioactivities remaining in the slices after superfusion and recovered in the collected superfusate fractions (Kontro and Oja, 1987). The efflux rate constants of taurine for the time intervals of 20 to $30 \, \text{min}$ (k_1) and 34 to $50 \, \text{min}$ (k_2) were computed as negative slopes for the regression lines of the logarithm of radioactivity remaining in the slices vs. superfusion time.

Experimental conditions

Neural cell damage was induced by modifying experimental conditions from the beginning of superfusions. The medium for metabolic blockade contained 1.0 mM NaCN or 1.0 mM 2,4-dinitrophenol (DNP). In hypoglycemia, glucose was omitted from the medium and in hypoxia the medium was bubbled with N_2 gas for 1h before and during the experiments. In ischemia, the glucose-free medium was bubbled with N_2 gas. Lipid peroxidation (Wills, 1969) (oxidative stress) was induced by FeSO₄ (7.5 μ M) together with ascorbate (1.5 mM) (Agostinho et al., 1994). Free radical production was achieved by exposure to 0.01% hydrogen peroxide (Gilman et al., 1994).

Statistical calculations

The presence of statistically significant differences between the sample means was detected by variance analysis. Comparisons of individual means were made by Hartley's sequential method of testing.

Results

The glutamate receptor agonists NMDA, kainate and AMPA (all 0.1 mM) significantly potentiated the release of [3H]taurine from hippocampal slices

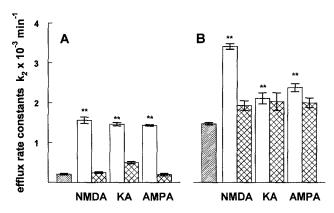


Fig. 1. Effects of ionotropic glutamate agonists (all 0.1 mM) NMDA, kainate and AMPA (open columns) on taurine release from hippocampal slices from 7-day- ($\bf A$) and 3-monthold ($\bf B$) mice in normoxic control conditions. The results are efflux rate constants (\pm SEM) k_2 for the time interval 34–50 min. The shaded columns depict control release without effectors. The cross-hatched columns represent the effects of NMDA, kainate and AMPA together with their respective antagonists MK-801, CNQX and NBQX (all 0.1 mM). The number of independent experiments is 4–8. Significance of differences from the control: **p < 0.01

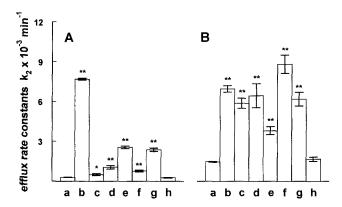


Fig. 2. Taurine release from hippocampal slices from 7-day- (**A**) and 3-month-old (**B**) mice in cell-damaging conditions. The results are mean efflux rate constants (\pm SEM) k₂ (34–50 min) of 4–8 independent experiments; (a) control release, and the effects of (b) 1.0 mM 2,4-dinitrophenol, (c) 1.0 mM NaCN, (d) hypoglycemia, (e) hypoxia, (f) ischemia, (g) free radicals and (h) oxidative stress. Significance of differences from the control: *p < 0.05, **p < 0.01

from both 7-day- and 3-month-old mice (Fig. 1). NMDA was the most potent at both ages. The effects of NMDA, kainate and AMPA were abolished by their antagonists MK-801, 6-cyano-7-nitroquinoxaline-2,3-dione (CNQX) and 6-nitro-7-sulphamoylbenzo[f]quinoxaline-2,3-dione (NBQX), respectively, only CNQX being unable to reduce the potentiation by kainate significantly in the adults (Fig. 1). All cell-damaging conditions, applied from the beginning

of superfusions, markedly enhanced taurine release, except for oxidative stress (Fig. 2). In the developing hippocampus 1.0 mM DNP was the most potent, followed by hypoxia and exposure to free radicals. Ischemia was the most effective in the adults, but also DNP, hypoglycemia, free radicals and NaCN (1.0 mM) evoked substantial taurine release.

The agonists NMDA, kainate and AMPA had no effect on taurine release in the presence of DNP (data not shown). In the presence of NaCN the agonists were also without effect in the adult hippocampus, but all three markedly potentiated taurine release in developing mice. These effects were abolished by the respective antagonists (Fig. 3). In hypoxic conditions the agonists were able to enhance taurine release in the immature hippocampus, the antagonists again blocking the effects (Fig. 4). In the adults kainate and AMPA were effective, their actions being reduced by CNQX and NBQX, respectively. In hypoglycemia and ischemia the agonists were unable to potentiate taurine release in the adult hippocampus, but in the immature hippocampus NMDA and kainate enhanced the release in hypoglycemia, which effects were blocked by MK-801 and CNQX, respectively (Fig. 5A, Fig. 6). Moreover, all agonists were effective in ischemic conditions in developing mice but the NMDA effect was not reduced by MK-801 (Fig. 5B). In freeradical-containing medium the agonists had no effects on taurine release, neither in the adult nor in the developing hippocampus (data not shown). On the other hand, all agonists potentiated the release under oxidative stress in both age groups. Their effects were antagonized by MK-801, CNQX and NBQX, except for the AMPA effect in the adults (Fig. 7).

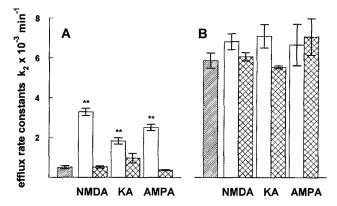


Fig. 3. Effects of NMDA, kainate and AMPA (open columns) (all 0.1 mM) on hippocampal taurine release in 7-day- (**A**) and 3-month-old (**B**) mice in the presence of 1.0 mM NaCN. The results are mean efflux rate constants (\pm SEM) k₂ (34–50 min) of 4–8 independent experiments. The shaded columns represent control release and the cross-hatched columns the effects of the agonists together with their respective antagonists MK-801, CNQX and AMPA (all 0.1 mM). Significance of differences from the control: **p < 0.01

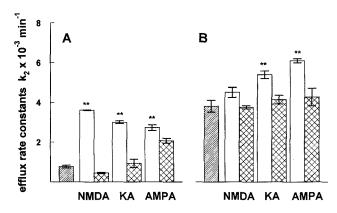


Fig. 4. Effects of NMDA, kainate and AMPA (open columns) (all 0.1 mM) on hippocampal taurine release in 7-day- (**A**) and 3-month-old (**B**) mice in hypoxic conditions. The results are mean efflux rate constants (\pm SEM) k₂ (34–50 min) of 4–8 independent experiments. The shaded columns represent control release and the cross-hatched columns the effects of the agonists together with their respective antagonists MK-801, CNQX and NBQX (all 0.1 mM). Significance of differences from the control: **p < 0.01

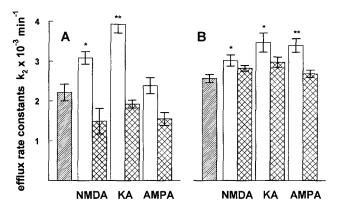


Fig. 5. Effects of NMDA, kainate and AMPA (open columns) (all $0.1\,\mathrm{mM}$) on hippocampal taurine release in 7-day-old mice in hypoglycemia (A) and ischemia (B). The results are mean efflux rate constants ($\pm \mathrm{SEM}$) k_2 (34–50 min) of 4–8 independent experiments. The shaded columns represent control release and the cross-hatched columns the effects of the agonists together with their respective antagonists MK-801, CNQX and NBQX (all $0.1\,\mathrm{mM}$). Significance of differences from the control: *p < 0.05, **p < 0.01

Discussion

The hippocampal release of taurine was modified in both adult and developing mice by ionotropic glutamate receptors. NMDA, kainate and AMPA all markedly potentiated taurine release in the developing hippocampus, whereas the effects were considerably smaller in the adults. NMDA has generally been reported to be the most powerful agonist (Magnusson et al., 1991; Saransaari and Oja, 1991; 1994). The effects of kainate and AMPA were

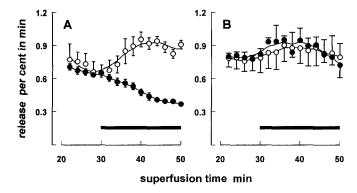


Fig. 6. Release of taurine from hippocampal slices from 7-day- (A) and 3-month-old (B) mice in hypoglycemia in the presence of 0.1 mM kainate (-o-) and 0.1 mM kainate together with 0.1 mM CNQX (-◆). The results are mean values ±SEM of 4 separate experiments. Kainate and CNQX were present in superfusion medium from 30 to 50 min, as indicated by the bar

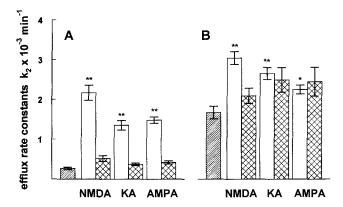


Fig. 7. Effects of NMDA, kainate and AMPA (open columns) (all 0.1 mM) on hippocampal taurine release in 7-day- (A) and 3-month-old (B) mice in medium inducing oxidative stress. The results are mean efflux rate constants (\pm SEM) k₂ (34–50 min) of 4–8 independent experiments. The shaded columns represent control release and the cross-hatched columns the effects of the agonists together with their respective antagonists MK-801, CNQX and NBQX (all 0.1 mM). Significance of differences from the control: *p < 0.05, **p < 0.01

abolished by the non-NMDA antagonists CNQX and NBQX, respectively, indicating that both types of receptors are involved in the evoked release of taurine in the immature hippocampus. The regulation by kainate receptors seems to be lost during maturation, since CNQX was unable to affect the kainate-evoked release in the adults. The inhibition of AMPA effects by NBQX implies the participation of AMPA receptors also in the adult hippocampus. The stimulation by NMDA was blocked by the specific antagonist MK-801 in both age groups. The occurrence of presynaptic NMDA-receptor-mediated release of taurine is corroborated by the finding that another potent NMDA receptor agonist, tetrazolylglycine (Schoepp et al.,

1991), has also enhanced taurine release in a dizocilpine-sensitive manner in both developing and adult mice (Saransaari and Oja, 1994). The operation of presynaptic glutamate receptors regulating the release of glutamate (Smirnova et al., 1993), together with that of other transmitters, including noradrenaline (Pittaluga and Raiteri, 1992a,b) and GABA (Janáky et al., 1993), has been demonstrated in the adult hippocampus. In the immature hippocampus, the agonists also potentiate GABA release in a receptor-mediated manner, implying the existence of presynaptic glutamate receptors regulating GABA release already during the early postnatal period (Saransaari and Oja, 1997b).

The ability of NMDA to evoke substantial release of taurine in the immature hippocampus tallies with the overexpression of the NMDA subtype of glutamate receptors in hippocampal formation during postnatal development (Tremblay et al., 1988; Represa et al., 1989; McDonald et al., 1990; Le Greves et al., 1996). The increase in the number of receptor sites also correlates with the development of afferent input and elaboration of dendrites in the hippocampus and with the developmental onset of long-term potentiation (Pokorny and Yamamoto, 1981a,b). During this period the number of kainate and AMPA binding sites is relatively low in the rat hippocampus, increasing only somewhat later (Ben-Ari et al., 1984; Insel et al., 1990; Miller et al., 1990). In rodents postsynaptic excitation consequently predominates over inhibition during the first weeks of life. This early developmental overexpression of NMDA receptors coincides with the increased susceptibility to seizures and excitotoxicity produced by excitatory amino acids (McDonald and Johnston, 1990). At this time the pronounced glutamate-receptor-activated release of taurine may be of great importance in counteracting excitotoxic effects and protecting against impending hyperexcitation.

Taurine is metabolically rather inert in the brain. The compound is not broken down (Huxtable, 1992) and the de novo synthesis is slow (Oja et al., 1973; Oja and Kontro, 1981). The blood-brain exchange rates are likewise low in both adult and developing rodent brain (Oja et al., 1976). The brain slices also maintain well their intracellular levels of taurine upon incubation in vitro, the slices from the developing brain in particular (Oja, 1971). The present qualitative differences in the responses to the glutamate receptor agonists and antagonists cannot be accounted for any differences in taurine metabolism in the two experimental groups. Although the taurine concentration in hippocampal slices is more than two times higher in 7-day-old than in 3month-old mice, the spontaneous release of endogenous taurine is threefold in the latter (Saransaari and Oja, unpublished results). The preloaded [3H]taurine is then mixed in adult mice with a smaller intracellular taurine pool that is more readily spontaneously releasable. The quantitative responses to the effectors are thus to some extent overshadowed by spontaneous taurine release in slices from the mature hippocampus.

The conditions known to cause neural cell damage (Haddad and Jiang, 1993; Hara et al., 1993) generally greatly enhanced taurine release. The marked increase in taurine release in cell-damaging conditions, particularly in ischemia, could be attributable to several mechanisms, including

Ca-dependent exocytosis, Ca-independent release via reversal of carriermediated uptake, indiscriminate opening of ion channels and unspecific leakage through damaged plasma membranes. This last alternative is not likely, however, since earlier investigations have shown that in hippocampal slices the liberation of lactate dehydrogenase (a common marker of plasma membrane damage and nonspecific lysis of neural cells) is not increased by the present type of ischemia (Pellegrini-Giampietro et al., 1990; Cherici et al., 1991). Neither could we now see any increase in lactate dehydrogenase in superfusion media under ischemic conditions in either age group (data not shown). An activation of volume-sensitive efflux could have contributed to taurine release (Schousboe et al., 1990). For example, brain slices swell under anaerobic conditions and in the presence of metabolic poisons (Laakso and Oja, 1976). Intracellular swelling activates stretch-sensitive ion channels and is generally accompanied by the release of both inorganic and organic osmolytes, including taurine (Pasantes-Morales et al., 1993). To date, nonsaturable diffusion of taurine is indeed known to be greatly enhanced in ischemic conditions (Saransaari and Oja, 1996).

The substantial increase in the release of taurine in ischemia and other cell-damaging conditions could be due partly to inhibition of reuptake. Taurine uptake by mouse cerebral cortical synaptosomes is in fact inhibited in most of the present cell-damaging conditions, most markedly in the adult brain (Saransaari and Oja, 1996). Although the medium is continuously renewed in superfusion experiments, taurine molecules released from the intracellular spaces must first traverse the extracellular space, being then subject to possible reuptake. Indeed, there is a significant positive correlation between the efflux and influx rates of taurine in brain slices in different experimental conditions (Oja and Saransaari, 1994). The enhanced release of excitatory amino acids in hypoxic and ischemic conditions (Lekieffre et al., 1992; Collard and Menon-Johansson, 1993; Globus et al., 1993) has also been thought to result from a reversed operation of cell membrane carriers (Szatkowski and Attwell 1994). The function of Na-dependent aturine carriers in cell plasma membranes is similarly reversible in certain experimental conditions (Korpi and Oja, 1983). The reversed action of sodium-dependent high-affinity taurine carriers, changing the direction of mediated transport, has thus apparently contributed to the present results.

The ability of ionotropic glutamate agonists to evoke taurine release varied in the different cell-damaging conditions. In the adults glutamate agonists were able to evoke release only in hypoxia and oxidative stress. Generally, the glutamate-receptor-stimulated release was operative in the immature hippocampus, except in the presence of DNP and free radicals, which agents as such already substantially potentiated the release. Free radicals are normal by-products of oxidative metabolism. Under certain conditions, including hypoxia and ischemia, free radical production may be excessively increased, damaging nervous system functions by disrupting membrane structures (Pelmar et al., 1989). Free radicals are also thought to be involved in many neurological disorders (Halliwell, 1992). Taurine release was greatly potentiated by exposure to media producing free radicals in both mature

and immature hippocampus, and the glutamate agonists failed to evoke any additional release. The release of glutamate has also been enhanced by exposure to peroxide-generated free radicals in rat hippocampal slices (Pellegrini-Giampietro et al., 1988, 1990), leading to extracellular accumulation of excitatory amino acids and contributing to excitotoxicity.

It is of particular note that in hypoxia, oxidative stress and metabolic blockade by NaCN, the activation of the three ionotropic glutamate receptors can enhance taurine release in the developing hippocampus. In ischemia the NMDA-induced release is apparently not receptor-mediated, since MK-801 was not able to antagonize it. Moreover, in hypoglycemia both NMDA and kainate receptors induced taurine release. This glutamate-receptor-evoked massive release of taurine could have significant neuroprotective effects in the immature hippocampus, countering in several ways the harmful actions of simultaneously liberated excitatory amino acids. Taurine inhibits the depolarizing effects of excitatory amino acids by increasing membrane chloride conductance (Oja et al., 1990). It attenuates Ca²⁺ influx in adult and developing brain tissue and antagonizes depolarization-evoked Ca²⁺ efflux (Kontro and Oja, 1988). Furthermore, intracellular swelling of neurons and glial cells due to activation of glutamate receptors (Saransaari and Oja, 1991) is also attenuated by the extracellularly released taurine (Walz and Allen, 1987).

Acknowledgements

The skillful technical assistance of Mrs Irma Rantamaa and Mrs Oili Pääkkönen and the financial support of the Medical Research Fund of Tampere University Hospital are gratefully acknowledged.

References

- Agostinho P, Duarte CB, Carvalho AP, Oliveira CR (1994) Effect of oxidative stress on the release of [3H]GABA in cultured chick retina cells. Brain Res 655: 213–221
- Ben-Ari Y, Tremblay E, Berger M, Nitecka L (1984) Kainic acid seizure syndrome and binding sites in developing rats. Dev Brain Res 14: 284–288
- Benveniste H, Grejer J, Schousboe A, Diemer NH (1984) Elevation of the extracellular concentrations of glutamate and aspartate in rat hippocampus during transient cerebral ischemia monitored by intracerebral microdialysis. J Neurochem 43: 1369–1374
- Cherici G, Alesiani M, Pellegrini-Giampiertro DE, Moroni F (1991) Ischemia does not induce the release of excitotoxic amino acids from the hippocampus of newborn rats. Dev Brain Res 60: 235–240
- Cherubini E, Ben-Ari Y, Krnjevic K (1989) Anoxia produces smaller changes in synaptic transmission, membrane potential and input resistance in immature rat hippocampus. J Neurophysiol 62: 882–895
- Choi DW, Rothman SM (1990) The role of glutamate neurotoxicity in hypoxic-ischemic neuronal death. Annu Rev Neurosci 13: 171–182
- Collard KJ, Menon-Johansson AS (1993) Effects of short-term hypoxia on [³H]glutamate release from preloaded hippocampal and cortical synaptosomes. Neurochem Res 8: 165–170
- Cook TM, Crutcher KA (1986) Intrahippocampal injection of kainic acid produces significant pyramidal cell loss in neonatal rats. Neuroscience 18: 79–92

- Coyle JT, Puttfarcken P (1993) Oxidative stress, glutamate and neurodegenerative disorders. Science 262: 689–695
- Ferriero DM, Arcavi LJ, Simon RP (1990) Ontogeny of excitotoxic injury to nicotinamide adenine dinucleotide phosphate diaphorase reactive neurons in the neonatal rat striatum. Brain Res 304: 417–424
- French ED, Vezzani A, Whetsell Jr WO, Schwarcz R (1986) Antiexcitotoxic actions of taurine in the rat hippocampus studied in vivo and in vitro. Adv Exp Med Biol 203: 349–362
- Gilman SC, Bonner MJ, Pellmar TC (1994) Free radicals enhance basal release of D-[³H]aspartate from cerebral cortical synaptosomes. J Neurochem 62: 1757–1763
- Globus MY-T, Busto R, Dietrich WD, Martinez E, Valdes I, Ginsberg MD (1988) Effect of ischemia on the in vivo release of striatal dopamine, glutamate and γ-aminobutyric acid studied by intracerebral microdialysis. J Neurochem 51: 1455–1464
- Haddad GG, Jiang C (1993) O_2 deprivation in the central nervous system: on mechanisms of neuronal response, differential sensitivity and injury. Prog Neurobiol 40: 277–318
- Hagberg H, Lehmann A, Sandberg M, Nyström B, Jacobson I, Hamberger A (1985) Ischemic-induced shifts of inhibitory and excitatory amino acids in area CA1 of the rat hippocampus. Dev Brain Res 38: 286–290
- Halliwell B (1992) Reactive oxygen species and the central nervous system. J Neurochem 59: 1609–1623
- Hara H, Sukamoto T, Kogure K (1993) Mechanism and pathogenesis of ischemia-induced neuronal damage. Prog Neurobiol 40: 645–670
- Huxtable RJ (1992) The physiological actions of taurine. Physiol Rev 72: 101–163
- Ikonomidou C, Price MT, Mosinger JL, Friedrich G, Labruyere J, Salles KS, Olney J (1989) Hypobaric-ischemic conditions produce glutamate-like cytopathology in infant rat brain. J Neurosci 9: 1693–1700
- Insel TR, Miller LP, Gelhard RE (1990) The ontogeny of excitatory amino acid receptors in rat forebrain. I. N-methyl-D-aspartate and quisqualate receptors. Neuroscience 35: 31–43
- Janáky R, Saransaari P, Oja SS (1993) Release of GABA from rat hippocampal slices: involvement of quisqualate/N-methyl-D-aspartate-gated ionohpores and extracellular magnesium. Neuroscience 53: 779–785
- Kontro P, Oja SS (1987) Taurine and GABA release from mouse cerebral cortex slices: potassium stimulation releases more taurine than GABA from developing brain. Dev Brain Res 37: 277–291
- Kontro P, Oja SS (1988) Effects of taurine on the influx and efflux of calcium in brain slices of adult and developing mice. Int J Neurosci 38: 103–109
- Kontro P, Marnela K-M, Oja SS (1980) Free amino acids in the synaptosome and synaptic vesicle fractions of different bovine brain areas. Brain Res 184: 129–141
- Korpi ER, Oja SS (1983) Characteristics of taurine release from cerebral cortex slices induced by sodium-deficient media. Brain Res 289: 197–204
- Laakso M-L, Oja SS (1976) Factors influencing the inulin space in cerebral cortex slices from adult and 7-day-old rats. Acta Physiol Scand 97: 486–494
- Le Grevés P, Hoogendoorn K, Synnergren B, Meyerson B, Nyberg F (1996) The relationship between the NMDA receptor NR1 subunit mRNA and [3H]MK-801 binding in the embryonic and early postnatal rat CNS. Neurosci Res Commun 19: 145–152
- Lekieffre D, Callebert J, Plotkine M, Boulu RG (1992) Concomitant increases in the extracellular concentrations of excitatory and inhibitory amino acids in the rat hippocampus during forebrain ischemia. Neurosci Lett 137: 78–82
- Lombardini JB (1976) Regional and subcellular studies on taurine in the rat central nervous system. In: Huxtable R, Barbeau A (eds) Taurine. Raven Press, New York, pp 311–326

- Magnusson KR, Clements JR, Wu J-Y, Beitz AJ (1989) Colocalization of taurine- and cysteine sulfinic acid decarboxylase-like immunoreactivity in the hippocampus of the rat. Synapse 4: 55–69
- Magnusson KR, Koerner JF, Larson AA, Smullin DH, Skilling SR, Beitz AJ (1991) NMDA-, kainate- and quisqualate-stimulated release of taurine from electrophysiologically monitored rat hippocampal slices. Brain Res 549: 1–8
- McDonald JW, Johnston MV (1990) Physiological and pathophysiological roles of excitatory amino acids during central nervous system development. Brain Res Rev 15: 41–70
- McDonald JW, Johnston MV, Young AB (1990) Differential ontogenic development of three receptors comprising the NMDA receptor/channel complex in the rat hippocampus. Exp Neurol 110: 237–247
- Miller LP, Johnson AE, Gelhard RE, Insel TR (1990) The ontogeny of excitatory amino acid receptors in the rat forebrain II. Kainic acid receptors. Neuroscience 35: 45–51
- Obrenovitch TP, Urenjak J (1997) Altered glutamatergic transmission in neurological disorders: from high extracellular glutamate to excessive synaptic efficacy. Prog Neurobiol 51: 39–87
- Oja SS (1971) Exchange of taurine in brain slices of adult and 7-day-old rats. J Neurochem 18: 1847–1852
- Oja SS, Kontro P (1981) Oxidation of hypotaurine in vitro by mouse liver and brain tissues. Biochim Biophys Acta 677: 350–357
- Oja SS, Kontro P (1983) Taurine. In: Lajtha A (ed) Handbook of neurochemistry, vol 3, 2nd edn. Plenum Press, New York, pp 501–533
- Oja SS, Karvonen M-L, Lähdesmäki P (1973) Biosynthesis of taurine and enhancement of decarboxylation of cysteine sulphinate and glutamate by the electrical stimulation of rat brain slices. Brain Res 55: 173–178
- Oja SS, Lehtinen I, Lähdesmäki P (1976) Taurine transport rates between plasma and tissues in adult and 7-day-old mice. Q J Exp Physiol 61: 133–143
- Oja SS, Korpi ER, Saransaari P (1990) Modification of chloride flux across brain membranes by inhibitory amino acids in developing and adult mice. Neurochem Res 15: 797–804
- O'Regan MH, Smith-Barbour M, Perkins LM, Phillis JW (1995) A possible role for phospholipases in the release of neurotransmitter amino acids from ischemic rat cerebral cortex. Neurosci Lett 185: 191–194
- Pasantes-Morales H, Alavez S, Sánchez Olea R, Morán J (1993) Contribution of organic and inorganic osmolytes to volume regulation in rat brain cells in culture. Neurochem Res 18: 445–452
- Pellmar TC, Neel KL, Lee KH (1989) Free radicals mediate peroxidative damage in the guinea pig hippocampus in vitro. J Neurosci Res 24: 437–444
- Pellegrini-Giampietro DE, Cherici G, Alesiani M, Carlá V, Moroni F (1988) Excitatory amino acid release from rat hippocampal slices as a consequence of free-radical formation. J Neurochem 51: 1960–1963
- Pellegrini-Giampietro DE, Cherici G, Alesiani, M, Carlá V, Moroni F (1990) Excitatory amino acid release and free radical formation may cooperate in the genesis of ischemia-induced neuronal damage. J Neurosci 10: 1035–1041
- Pittaluga A, Raiteri M (1992a) N-Methyl-D-aspartic acid (NMDA) and non-NMDA receptors regulating hippocampal norepinephrine release. III. Changes in the NMDA receptor complex induced by their functional cooperation. J Pharmacol Exp Ther 263: 327–333
- Pittaluga A, Raiteri M (1992b) N-Methyl-D-aspartic acid (NMDA) and non-NMDA receptors regulating hippocampal norepinephrine release. I. Location on axon terminals and pharmacological characterization. J Pharmacol Exp Ther 260: 232–237
- Pokorny J, Yamamoto T (1981a) Postnatal ontogenesis of hippocampal CA1 area in rats. I. Development of dendritic arborization in pyramidal neurons. Brain Res Bull 7: 113–120

- Pokorny J, Yamamoto T (1981b) Postnatal ontogenesis of hippocampal CA1 area in rats. II. Development of ultrastructure in stratum lacunosum and moleculare. Brain Res Bull 7: 121–130
- Pulsinelli WA, Brierley JB, Plum F (1982) Temporal profile of neuronal damage in a model of transient forebrain ischemia. Ann Neurol 11: 491–498
- Represa A, Tremblay E, Ben-Ari Y (1989) Transient increase of NMDA-binding sites in human hippocampus during development. Neurosci Lett 99: 61–66
- Rothman SM, Olney JW (1988) Glutamate and pathology of hypoxic/ischemic brain damage. Ann Neurol 19: 105–111
- Saransaari P, Oja SS (1991) Excitatory amino acids evoke taurine release from cerebral cortex slices from adult and developing mice. Neuroscience 45: 451–459
- Saransaari P, Oja SS (1992) Taurine transport in the mouse cerebral cortex during development and ageing. Adv Exp Med Biol 315: 215–220
- Saransaari P, Oja SS (1994) Taurine release from mouse hippocampal slices: effects of glutamatergic substances and hypoxia. Adv Exp Med Biol 359: 279–287
- Saransaari P, Oja SS (1996) Taurine and neural cell damage: transport of taurine in adult and developing mice. Adv Exp Med Biol 403: 481–490
- Saransaari P, Oja SS (1997a) Enhanced taurine release in cell-damaging conditions in the developing and ageing mouse hippocampus. Neuroscience 79: 847–854
- Saransaari P, Oja SS (1997b) Enhanced GABA release in cell-damaging conditions in the adult and developing mouse hippocampus. Int J Dev Neurosci 15: 163–174
- Schoepp DD, Smith CL, Lodge D, Millar JD, Leander JD, Sacaan AI, Lunn WHW (1991) D,L-(Tetrazol-5-yl)glycine: a novel and highly potent NMDA receptor agonist. Eur J Pharmacol 203: 237–243
- Schousboe A, Morán J, Pasantes-Morales H (1990) Potassium-stimulated release of taurine from cultured cerebellar granule cells is associated with cell swelling. J Neurosci Res 27: 71–77
- Schurr A, Tseng MT, West CA, Rigor BM (1987) Taurine improves the recovery of neuronal function following cerebral hypoxia: an in vitro study. Life Sci 40: 2059–2066
- Smirnova T, Stinnakre J, Mallet J (1993) Characterization of a presynaptic glutamate receptor. Science 262: 430–433
- Sturman JA (1993) Taurine in development. Physiol Rev 73: 119–147
- Taber KH, Lin C-T, Liu J-W, Thalmann R, Wu J-Y (1986) Taurine in hippocampus: localization and postsynaptic action. Brain Res 386: 113–121
- Tang XW, Deupree DL, Sun Y, Wu J-Y (1996) Biphasic effect of taurine on excitatory amino acid-induced neurotoxicity. Adv Exp Med Biol 403: 499–505
- Tremblay E, Roisin MP, Represa A, Charriant-Marlangue C, Ben-Ari Y (1988) Transient increased density of NMDA binding sites in the developing rat hippocampus. Brain Res 461: 393–396
- Trenkner E (1990) The role of taurine and glutamate during early postnatal cerebellar development of normal and weaver mutant mice. Adv Exp Med Biol 268: 239–244
- Walz W, Allen AF (1987) Evaluation of the osmoregulatory function of taurine in brain cells. Exp Brain Res 68: 290–298
- Wills ED (1969) Lipid peroxide formation in microsomes. General considerations. Biochem J 113: 315–324
- Wu J-Y, Lin C-T, Johanssen FF, Liu J-W (1994) Taurine neurons in rat hippocampal formation are relatively inert to cerebral ischemia. Adv Exp Med Biol 359: 289–298
- Zorumski CF, Olney JW (1993) Excitotoxic neuronal damage and neuropsychiatric disorders. Pharmacol Ther 59: 145–162

Authors' address: Prof. P. Saransaari, Tampere Brain Research Center, University of Tampere Medical School, Box 607, FIN-33101 Tampere, Finland.